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Reaction of advanced inbred lines of Habanero pepper to *Ralstonia pseudosolanacearum* and *Phytophthora capsici*

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ABSTRACT

Habanero pepper is important in the international market and is becoming popular in the national market; however, few cultivars adapted and resistant to devastating diseases caused by the soilborne pathogens *Ralstonia pseudosolanacearum* (RP) and *Phytophthora capsici* (PC) are available in Brazil. The aim of this study was to evaluate the reaction of Habanero-type advanced inbred lines, developed by Embrapa Hortaliças breeding program, to RP and PC. Although not required for the process of protection, registration and release of new cultivars, this information is important. CNPH 15.737; CNPH 15.740; CNPH 15.744; CNPH 15.745; CNPH 15.749 and CNPH 15.750 were inoculated with three RP and one PC isolates. Plants showing wilted leaves (RP) and stem necrosis, leaf wilting and damping off (PC) were quantified. Total area under the disease progress curve (AUDPC) was obtained using incidence values and also severity index for RP. CNPH 15.740 and CNPH 15.737 were highly resistant to RP. CNPH 15.749 displayed considerable resistance levels to PC. CNPH 15.740 showed high resistance to RP isolates and intermediate resistance to PC isolates which also shows agronomic traits of interest to the sector, therefore with a high potential to be released as a new cultivar.

Keywords: *Capsicum*, bacterial wilt, *Phytophthora* wilt, incidence, severity index, resistance.

RESUMO

Reação de linhagens avançadas de pimenta Habanero a *Ralstonia pseudosolanacearum* e *Phytophthora capsici*

A pimenta Habanero é popular no mercado internacional e começa a despertar maior interesse no nacional, porém poucas cultivares adaptadas às condições edafoclimáticas e com resistência às principais doenças como murcha bacteriana (MB) e murcha de fitófтора (MF), causadas respectivamente por *Ralstonia pseudosolanacearum* (RP) e *Phytophthora capsici* (PC), estão disponíveis no Brasil. O objetivo deste estudo foi avaliar a reação de linhagens avançadas de pimenta do grupo Habanero desenvolvidas pelo programa de melhoramento da Embrapa Hortaliças à MB e MF. Apesar de não ser exigida no processo de registro, proteção e lançamento de novas cultivares, esta informação é importante para os produtores. As linhagens avançadas identificadas como CNPH 15.737, CNPH 15.740, CNPH 15.744, CNPH 15.745, CNPH 15.749 e CNPH 15.750 foram inoculadas com três isolados de RP e um de PC. Quantificaram-se as plantas com folhas murchas (RP) e com necrose do caule, murcha das folhas e tombamento para PC. A área total abaixo da curva de progresso da doença (AACPD) foi obtida utilizando-se valores de incidência e, também, o índice de severidade para RP. CNPH 15.740 e CNPH 15.737 apresentaram alta resistência a RP. CNPH 15.749 apresentou nível moderado de resistência à PC. Merece destaque especial a linhagem CNPH 15.740, com nível elevado de resistência aos isolados de RP e intermediário a PC e que também possui características agrônomicas desejáveis tendo, portanto, alto potencial como nova cultivar a ser disponibilizada aos produtores.

Palavras-chave: *Capsicum*, murcha bacteriana, murcha de fitófтора, incidência, índice de severidade, resistência.

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Habanero is a highly diversified pepper group belonging to the species *Capsicum chinense*. Fruits can present different pungency levels, sizes and colors (Castro-Concha *et al.*, 2014). This group of pepper is mainly cultivated in tropical and subtropical regions (Teodoro *et al.*, 2013), especially in the

region of Yucatán, in Mexico (Bosland & Coon, 2015), where productivity can reach 40 t ha⁻¹ (Vázquez & Cortez, 2018).

Although originated in the Amazon basin, Habanero pepper is still little known in Brazil (Ribeiro *et al.*, 2015). However, interest on this pepper group

has been increasing at both national and international levels for the sauce and mash markets. This is due mainly to its unique flavor and aroma, besides generally high pungency levels (Nass *et al.*, 2015). Few Habanero-type cultivars are registered and adapted to Brazilian conditions. Thus, it is essential

to develop new cultivars adapted to Brazilian conditions, showing traits of interest such as pungency, outstanding flavor, strong aroma and high yield, as well as resistance to its main pathogens.

Among the pathogens that attack pepper plants (*Capsicum* spp.), stand out those that are soilborne and cause direct damages to the root system and to the stem base, consequently affecting the vegetative development and fruit production (Naresh *et al.*, 2019). The main soilborne pathogens of pepper plants reported are *Ralstonia pseudosolanacearum* sp. (RP; Lebeau *et al.*, 2011; Granke *et al.*, 2012; Rossato *et al.*, 2018), *Phytophthora capsici* (PC; Gilardi *et al.*, 2013; Reeves *et al.*, 2013; Sánchez-Chávez *et al.*, 2017), presented in this study, and *Meloidogyne* nematodes (Pinheiro *et al.*, 2014), which cause bacterial wilt, phytophthora blight and root knot, respectively.

The bacterium *Ralstonia pseudosolanacearum* originates from the recent taxonomic reorganization of *R. solanacearum*, previously identified as race 1 biovar 3, phylotype I (Rossato *et al.*, 2018). This species is more aggressive than *R. solanacearum* (races 1, 2, 3, biovars 1, 2, phylotype II) on *Capsicum*, and presents higher evolution rates and the largest geographical distribution worldwide (Lopes & Boiteux, 2004; Rossato *et al.*, 2018). This fact was also noticed under natural conditions in the Amazon region (Coelho Netto *et al.*, 2004). The bacterium penetrates the root system and proliferates in the xylem, causing irreversible wilting and, consequently, plant death (Lebeau *et al.*, 2011). Hong *et al.* (2012) reported that bacterial wilt affects solanaceous yield in over 80 countries, with annual losses of more than one billion dollars.

The oomycete *Phytophthora capsici* is found in practically all solanaceous-producing regions of the world and is considered one of the most destructive soilborne pathogens of *Capsicum* peppers (Sánchez-Chávez *et al.*, 2017). This phytopathogen causes root rot and stem base rot, leading to sudden wilting, dark brown necrosis of stem base and plant death (Dunn *et al.*, 2014). The variability of the species is represented

by more than 45 physiological races (Barchenger *et al.*, 2018). In Brazil, Ribeiro & Bosland (2012) identified eight physiological races of *P. capsici* in sweet pepper areas, and race 18 was the most common in the Central region. *P. capsici* reproduces both sexually and asexually, being characterized as a persistent problem, especially after consecutive cultivations of susceptible host plants (Naresh *et al.*, 2019). Its establishment and propagation are favored by excessive humidity, poor drainage and high soil temperature (Granke *et al.*, 2012; Petry *et al.*, 2016).

The use of resistant cultivars is considered the best management strategy to control diseases caused by *R. pseudosolanacearum* and *P. capsici* (Naresh *et al.*, 2019). Resistant cultivars are attractive since they are easy to be adopted by farmers and result in less environmental impacts than any other disease control strategy (Granke *et al.*, 2012). Sources of resistance to these two diseases were identified in accessions of *Capsicum* spp. (Madeira *et al.*, 2016; Petry *et al.*, 2016, Rossato *et al.*, 2018). Currently, the hybrid rootstock BRS Acará developed by Embrapa Hortaliças, has shown multiple resistance to several isolates of *P. capsici* and *R. pseudosolanacearum*, besides *Meloidogyne incognita* (Madeira *et al.*, 2016).

Little emphasis has been given to developing disease resistant cultivars of *Capsicum* spp., such as *C. baccatum*, *C. frutescens* and *C. chinense*. Embrapa Hortaliças' *Capsicum* breeding program aims to develop Habanero pepper genotypes resistant to *P. capsici* and *R. pseudosolanacearum*, as well as to other pathogens which attack the crop, besides attributes such as high yield, nutritional quality, and adaptation to tropical growing conditions. In this breeding program, six new advanced inbred lines (S₃), from a base population of Habanero pepper with a wide genetic variability (Nass *et al.*, 2015) were obtained after generations of selection using self-pollination methodology and individual plant selection with progeny test. These inbred lines are in the evaluation step for cultivar registration and protection of cultivars, mandatory by Brazilian

legislation.

The aim of this study was to evaluate the reaction of six advanced inbred lines of Habanero pepper from Embrapa Hortaliças' breeding program to *R. pseudosolanacearum* and *P. capsici*. Although not required, this information is important as additional data in the process of protection, registration and release of new cultivars.

MATERIAL AND METHODS

Genotypes and pathogens

Advanced inbred lines of Habanero pepper CNPH 15.737, CNPH 15.740, CNPH 15.744, CNPH 15.745, CNPH 15.749 and CNPH 15.750 (Table 1), from Embrapa Hortaliças' breeding program, were evaluated for reaction to *R. pseudosolanacearum* and *P. capsici*. Two experiments were carried out in a greenhouse of Embrapa Hortaliças, located in Brasília, Distrito Federal, Brazil, (15°55'60''W and 48°08'34''S, altitude 990 m), from November to December, 2018 (*P. capsici*) and from April to May, 2019 (*R. pseudosolanacearum*).

Reaction to inoculation with *R. pseudosolanacearum* isolates

The reaction of the six Habanero inbred lines to *R. pseudosolanacearum* was compared with two *C. annuum* genotypes used as control, CNPH 143 (resistant) and Tico (susceptible). Inbred lines and controls were sown in expanded polystyrene trays filled with sterile commercial substrate Carolina Soil® and kept in the greenhouse until inoculation and transplanting.

R. pseudosolanacearum isolates CNPH RS476, CNPH RS634 and CNPH RS639 were used in the inoculation. They were previously selected from the phytopathogenic bacteria collection of Embrapa Hortaliças for being highly virulent to pepper plants, as well as for the geographical diversity among isolates. These isolates were collected in different regions of Brazil, in Dom Pedro-MA, Guadalupe-PI and Altamira-PA, respectively. Inocula were prepared using the methodology described by Rossato *et al.* (2018).

At 45 days after sowing (DAS), seedlings of inbred lines and control genotypes were inoculated with the three *R. pseudosolanacearum* isolates, separately, by spraying 5 mL of a bacterial suspension, containing approximately 10^8 CFU mL⁻¹, directly on each plant root system exposed after removal from the tray (Lopes & Boiteux, 2004). After inoculation, the plants were transplanted to 1-liter plastic pots containing sterile soil and kept in a greenhouse during 27 days with night heating, preventing temperature drop below 20°C, which could increase the chance of escape. The temperature observed during the experimental period was 30±10°C.

Disease incidence was evaluated every three days. The evaluation started when the early symptoms appeared in the susceptible control [nine days after inoculation (DAI)]. Plants which presented wilting or the ones which died were counted, totalizing eight evaluations during the crop cycle (6, 9, 12, 15, 18, 21, 24 and 27 DAI), always in the afternoon in order to uniform the analysis. The last evaluation (at 27 DAI) occurred when the number of wilted plants stabilized.

Values of average disease incidence of each plot were used to calculate area under the disease progress curve (AUDPC) (Shaner & Finney, 1977). Disease severity was also evaluated at 26 DAI using a note scale from 1 to 5 (Winstead & Kelman, 1952), in which the lowest note corresponded to lack of wilting and the highest note to plant death. Notes 3 and above were attributed to plants which showed irreversible symptoms of wilting (susceptible) and notes between 1.5 to 2.5 were attributed to plants which showed light wilting symptoms, reversible after irrigation or at the coolest hours of the day.

A randomized complete block design, in factorial scheme 8 x 3 (six advanced inbred lines of Habanero and two controls CNPH 143 and Tico, and three *R. pseudosolanacearum* isolates), with four replicates, were used. Plots were composed of six plants, with two plants per pot.

Reaction to inoculation with *P. capsici* (isolate Pcp116, race 18)

Reactions of the six advanced Habanero lines to *P. capsici* were compared to those of *C. annuum* controls CNPH 148 (resistant) and bell pepper cultivar Ikeda (susceptible). Seedlings of inbred lines and control genotypes were produced according to what was described for the *R. pseudosolanacearum* experiment. At 45 DAS, seedlings were transplanted to 3-L plastic pots containing sterile soil. The base of each plant received 3 mL of a solution containing a suspension of 2×10^4 zoospores/mL⁻¹ of the Pcp 116 isolate, two days after transplant. The isolate was collected in the state of Goiás in 2007 and selected because of its high virulence. Inoculum was prepared based on the methodology described by Petry *et al.* (2016).

The experimental design was completely randomized, with eight treatments (inbred lines and controls) and five replications, three plants per experimental plot, totalizing 15 plants/treatment.

Assessment of disease incidence began at six DAI when first wilted plants of susceptible control (cultivar Ikeda) were observed. The evaluation was repeated every two days, totalizing six readings during the crop cycle. The incidence was evaluated using the quantity of plants with wilting symptoms, damping off and stem base necrosis in each experimental plot. The values of the average incidence of each plot were used to calculate the AUDPC (Shaner & Finney, 1977).

Statistical analysis

Variance of severity caused by RP and AUDPC for RP and PC were estimated. Homogeneity and normality

of residue of the mathematical model were tested, using Bartlett (Steel *et al.*, 1997) and Jarque-Bera (1987) tests, respectively. ANOVA with F test was carried out and the means among treatments were compared by Tukey test. In all statistical analyses, $p < 0.05$ was adopted. Statistical analyses were carried out using SISVAR v.5.6 statistical software (Ferreira, 2011), as well as Pearson correlation for AUDPC values and severity caused by RP.

RESULTS AND DISCUSSION

Reaction of inbred lines to *R. pseudosolanacearum*

Significant effect among Habanero pepper lines and *R. pseudosolanacearum* isolates was detected for both AUDPC and a single reading of disease severity at 26 DAI. There was significance for interaction between genotypes and isolates for both evaluations. Significant correlation was also observed between AUDPC and severity, with R² values for CNPH RS476, CNPH RS634 and CNPH RS639 isolates of 0.80; 0.87 and 0.88, respectively. Thus, it is possible to state that AUDPC and severity were highly associated parameters.

Inoculation was successful for all RP isolates and environmental conditions were favorable for disease development, allowing the differentiation of incidence and resistance classes among genotypes. The disease progressed rapidly in the susceptible control, as expected, showing first wilting symptoms at five DAI. The standard resistance control, CNPH 143, was asymptomatic to all isolates, as previously observed by

Table 1. Main plant and fruit traits of six Habanero pepper lines. Brasília, Embrapa Hortaliças, 2019.

Inbred lines	Plant architecture	Fruit shape	Fruit color	Pungency	Aroma
CNPH 15.737	Intermediate	Campanulated	Red	High	Medium
CNPH 15.740	Intermediate	Campanulated	Red	High	Medium
CNPH 15.744	Upright	Rectangular	Yellow	Low	Strong
CNPH 15.745	Intermediate	Campanulated	Red	Intermediate	Weak
CNPH 15.749	Intermediate	Campanulated	Red	Very high	Medium
CNPH 15.750	Intermediate	Campanulated	Red	Very High	Medium

Lopes & Boiteux (2004).

CNPH 15.737 and CNPH 15.740 displayed the lowest incidence, AUDPC and severity, in average values, for the three *R. pseudosolanacearum* isolates, differing significantly from the susceptible control (Figure 1, Table 2). CNPH 15.740 did not show wilted plants during the experiment when inoculated with isolate CNPH RS476. CNPH 15.737; CNPH 15.740 and CNPH 15.744 were highly resistant

to isolate CNPH RS476, not differing significantly from the resistant control CNPH 143 for all evaluations (Figure 1, Table 2). High resistance of inbred lines CNPH 15.737; CNPH 15.740 and CNPH 15.744 was also observed for isolate CNPH RS639, for the variables AUDPC and severity (Table 2). CNPH 15.737 did not differ significantly from CNPH 143 in severity when inoculated with isolate CNPH RS634. Although the inbred line CNPH 15.740 differed

significantly from the resistant control (CNPH 143) for CNPH RS634 isolate, for both AUDPC and severity, this inbred line presented a good level of resistance when compared to the susceptible control (Tico).

Different resistance levels of the evaluated inbred lines were observed in relation to *R. pseudosolanacearum* isolates. Inbred lines showed lower incidence of symptomatic plants for isolate CNPH RS476 in comparison to the highly virulent isolate CNPH RS634 (Figure 1). This fact reinforces the statement by Lopes *et al.* (2015) that the resistance to bacterial wilt is isolate specific rather than phylotype or biovar specific. This information should be taken into account in breeding programs aimed at resistance to bacterial wilt in different host plant species (Lopes & Boiteux, 2004; Wicker *et al.*, 2007).

Embrapa Hortaliças has been carrying out studies on identification and selection of resistance sources to *R. pseudosolanacearum* in *Capsicum* (Lopes & Boiteux, 2004; Rossato *et al.*, 2018) including the accessions CNPH 143 ('MC-4'), 'PBC 631', 'PBC 066', 'PBC 1347' and 'PBC 473' (Lopes & Boiteux, 2004) and CNPH 3800 (Rossato *et al.*, 2018). Cultivars of Habanero pepper BRS Nandaia and BRS Juruti, released previously by this breeding program, showed intermediate resistance to *R. pseudosolanacearum* (Ribeiro *et al.*, 2015).

Among the inbred lines studied, CNPH 15.740 and CNPH 15.737 stood out for presenting high and moderate resistance levels, respectively, to the three *R. pseudosolanacearum* isolates tested. CNPH 15.744 also stood out for presenting moderate resistance to isolates CNPH RS476 and CNPH RS639.

Reaction of inbred lines to *Phytophthora capsici*

The viability of the Pcp 116 isolate and the experimental conditions were satisfactory, since high incidence and AUDPC values for wilting, damping off and necrosis were observed in the cultivar Ikeda, used as susceptible control (Figure 2, Table 3). Ribeiro & Bosland (2012) reported the high

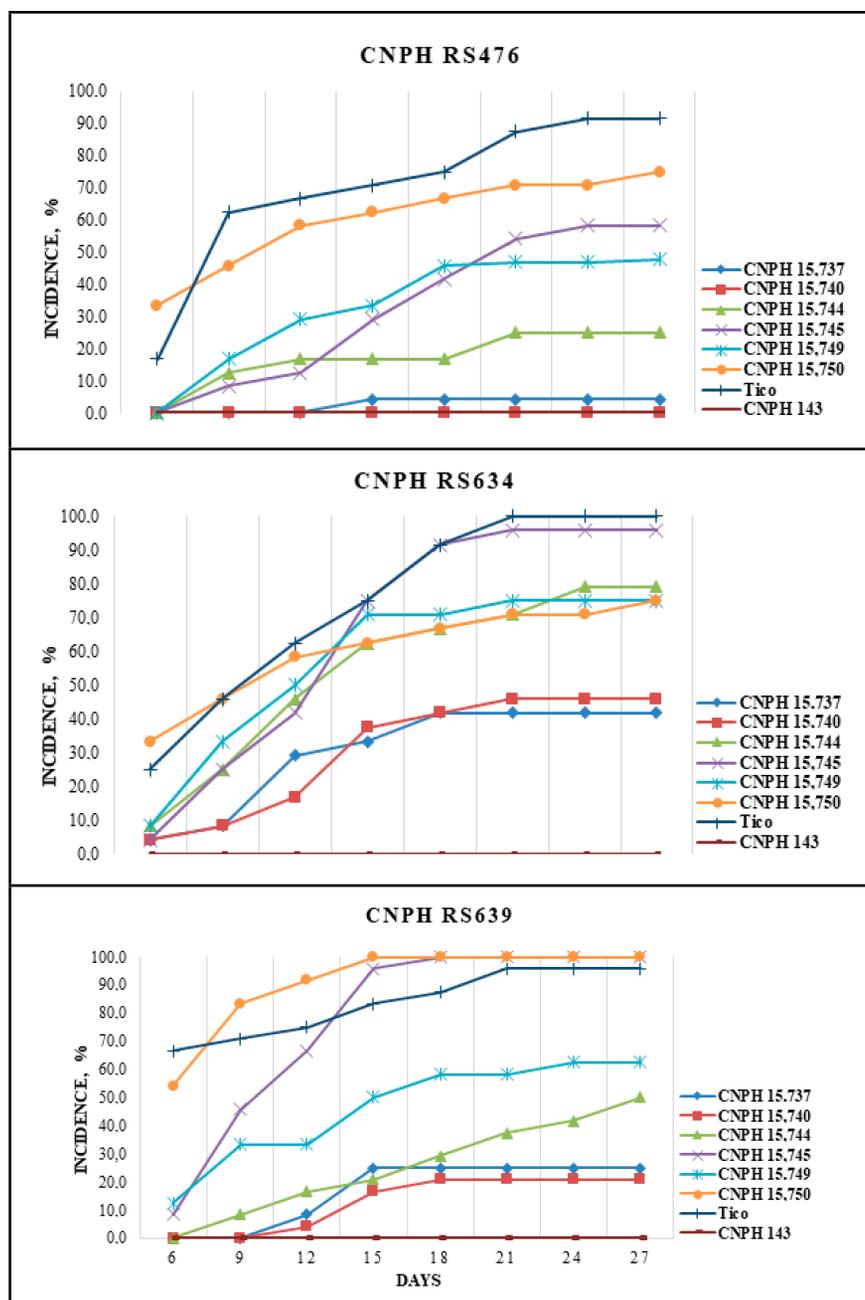


Figure 1. Plant wilt incidence in six Habanero pepper advanced inbred lines and susceptible (Tico) and resistant (CNPH 143) controls inoculated with three *Ralstonia pseudosolanacearum* isolates (CNPH RS476, CNPH RS634 and CNPH RS639). Brasília, Embrapa Hortaliças, 2019.

Table 2. Total area under the disease progress curve (AUDPC) and average values of severity of six Habanero pepper advanced inbred lines and in susceptibility (Tico) and resistance (CNPH 143) controls inoculated with three isolates of *Ralstonia pseudosolanacearum* (CNPH RS476, CNPH RS634 and CNPH RS639). Brasília, Embrapa Hortaliças, 2019.

Genotypes	CNPH RS476		CNPH RS634		CNPH RS639	
	AUDPC	Severity	AUDPC	Severity	AUDPC	Severity
CNPH 15.737	0.6 Cb ¹	1.1 BCa	5.9 Ca	1.6 DEa	3.3 Cab	1.4 DEa
CNPH 15.740	0 Cb	1.0 Cb	6.0 Ca	1.8 CDa	2.4 Cab	1.2 Eb
CNPH 15.744	3.3 BCb	1.2 BCb	10.5 BCa	2.4 BCa	4.8 BCb	1.3 Eb
CNPH 15.745	6.3 Bb	1.5 BCb	12.9 ABa	2.7 Ba	15.1 Aa	2.9 Ba
CNPH 15.749	6.3 Bb	1.8 ABb	11.1 Ba	2.7 Ba	8.8 Bab	2.0 CDb
CNPH 15.750	11.2 Ab	2.3 Ab	16.9 Aa	4.3 Aa	17.0 Aa	4.1 Aa
Tico	13.3 Aa	1.8 ABb	14.3 ABa	4.3 Aa	15.3 Aa	4.5 Aa
CNPH 143	0.0 Ca	1.0 Ca	0.0 Da	1.0 Ea	0.0 Ca	1.0 Ea
CV (%)	15.3	6.5	12.1	8.8	22.6	8.2

¹Average followed by same uppercase letters in the column and lowercase letters in the line does not differ significantly among the genotypes ($p < 0.05$) by Tukey test.

Table 3. Total area under the disease progress curve (AUDPC) for damping off, plant wilt and stem base necrosis in six Habanero pepper advanced lines and in susceptible (Ikeda) and resistant (CNPH 148) controls inoculated with *Phytophthora capsici* isolate Pcp 116. Brasília, Embrapa Hortaliças, 2019.

Genotypes	Damping off	Wilting	Necrosis
CNPH 15.737	0.97 c ¹	7.23 c	14.50 a
CNPH 15.740	0.40 c	6.30 c	12.13 b
CNPH 15.744	2.53 b	11.80 b	15.10 a
CNPH 15.745	0.80 c	8.17 bc	11.97 b
CNPH 15.749	0.00 c	1.17 de	11.10 b
CNPH 15.750	0.80 c	8.77 bc	11.93 b
Ikeda	13.50 a	15.30 a	15.20 a
CNPH 148	0.00 c	0.00 e	0.00 c
CV (%)	25.5	18.7	11.2

¹Average followed by same lowercase letter in the column does not differ significantly among the genotypes ($p < 0.05$) by Tukey test.

virulence of *P. capsici* race 18 based on pathogenicity reaction in pepper genotypes. The high virulence of this isolate was also verified in this study, considering that only the resistant control (CNPH 148) did not show symptomatic plants (Table 3).

All inbred lines evaluated, with exception of CNPH 15.744, showed higher levels of resistance to isolate Pcp116 than the susceptible control (Figure 2, Table 3). CNPH 15.749 presented the lowest AUDPC values for wilted plants, not differing significantly from the resistance control CNPH 148. For damping off, only the line

CNPH 15.744 was susceptible. CNPH 15.749; CNPH 15.740; CNPH 15.745 and CNPH 15.750 showed the lowest values of stem-base necrosis among the evaluated inbred lines, differing significantly from the susceptible control. However, the four inbred lines showed higher necrosis occurrence on the stem base in relation to the resistance control CNPH 148.

Infection by *P. capsici* may result in an expression of multiple disease symptoms, such as root rot, leaf wilt and stem necrosis, and each of these symptoms has a different resistance mechanism, requiring the presence of

specific resistance genes to each one (Reeves *et al.*, 2013; Barchenger *et al.*, 2017). Therefore, the occurrence of stem base necrosis in the studied inbred lines may be related to the presence of genes that differ from those that determine partial resistance observed in most of the studied inbred lines, taking into consideration wilting and damping off symptoms (Figure 2, Table 3).

Steiner & Bosland (2008) report that breeding for resistance to *P. capsici* in peppers is difficult and complex mainly due to the quantitative nature of inheritance (Naresh *et al.*, 2019). Naresh *et al.* (2019) highlight that the CM334 resistance (from the one which CNPH 148 was derived) to several *P. capsici* isolates is polygenic with additive and epistatic effect, so little success has been obtained in breeding programs using this genotype. The partial resistance of Habanero pepper lines in this study may be related to this complex effect, but further studies are necessary for its elucidation.

CNPH 15.749 stood out for presenting higher levels of resistance than other evaluated inbred lines and susceptible control, differing from resistance control CNPH 148 only in relation to the occurrence of stem-base necrosis. CNPH 15.740 showed moderate resistance to the *P. capsici* isolate.

The use of resistant cultivars

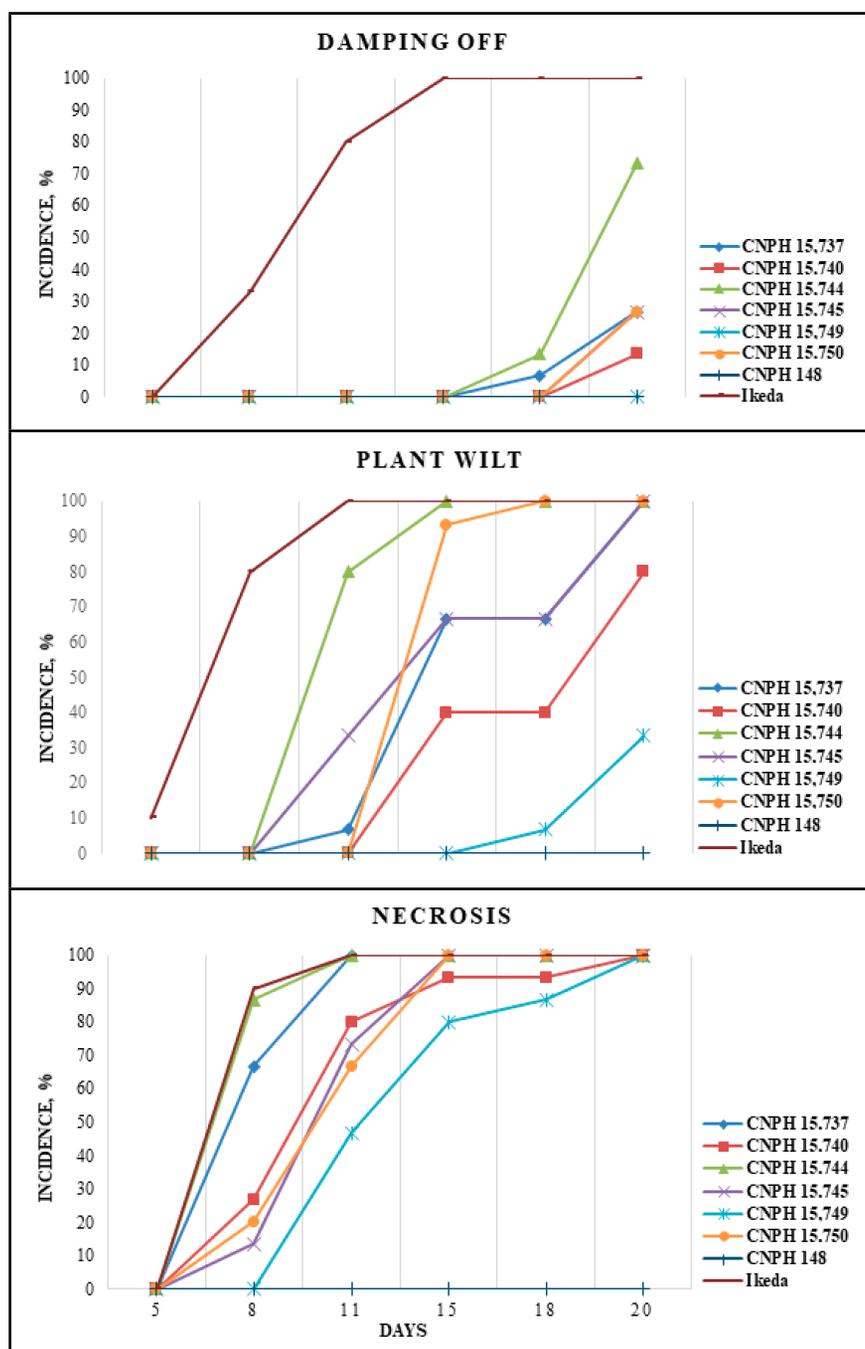


Figure 2. Damping off, plant wilt and stem base necrosis incidence in six Habanero pepper advanced lines and in susceptible (Ikeda) and resistant (CNPH 148) controls inoculated with *Phytophthora capsici* isolate Pcp 116. Brasília, Embrapa Hortaliças, 2019.

is the most suitable way to control diseases due to its low cost, high efficiency and reduced environmental impact, especially when compared to other strategies that aim to control the pathogen after its establishment (Rossato et al., 2018).

This study identified Habanero pepper inbred lines with high and intermediate levels of resistance to *R. pseudosolanacearum* and to *P. capsici*.

CNPH 15.737 and CNPH 15.740 were highly resistant to RP isolates and CNPH 15.749 presented considerable resistance *P. capsici* isolate Pcp 116. CNPH 15.740 stands out presenting a high level of resistance to RP isolates and intermediate resistance to PC; this line also presents desirable agronomic traits, such as high pungency, attractive flavor and aroma, intermediate plant architecture, important nutraceutical

compounds and high yield. The information obtained in this study will serve as complementary data to SNPC/MAPA technical forms for the registration and protection of cultivars in Brazil (National Cultivation Protection Service/Ministry of Agriculture, Livestock and Food Supply and RNC/MAPA - National Register of Cultivars).

REFERENCES

- BARCHENGER, DW; LAMOUR, KH; SHEU, Z; SHRESTHA, S; KUMAR, S; LIN, S; BURLAKOTI, R; BOSLAND, PW. 2017. Intra- and intergenomic variation of ploidy and clonality characterize *Phytophthora capsici* on *Capsicum* sp. in Taiwan. *Mycological Progress* 16: 955-963.
- BARCHENGER, DW; LAMOUR, KH; BOSLAND, PW. 2018. Challenges and strategies for breeding resistance in *Capsicum annuum* to the multifarious pathogen, *Phytophthora capsici*. *Frontiers in Plant Science* 9: 1-16.
- BOSLAND, PW; COON, D. 2015. 'NuMex Trick-or-Treat', a no-heat Habanero pepper. *HortScience* 50: 1739-1740.
- CASTRO-CONCHA, LA; TUYUB-CHE, J; MOO-MUKUL, A; VAZQUEZ-FLOTA, FA; MIRANDA-HAM, ML. 2014. Antioxidant capacity and total phenolic content in fruit tissues from accessions of *Capsicum chinense* Jacq. (Habanero pepper) at different stages of ripening. *The Scientific World Journal* 195: 1-5.
- COELHO NETTO, RA; PEREIRA, BG; NODA, H; BOHER, B. 2004. Murcha bacteriana no Estado do Amazonas, Brasil. *Fitopatologia Brasileira* 29: 21-27.
- DUNN, AR; LANGE, HW; SMART, CD. 2014. Evaluation of commercial bell pepper cultivars for resistance to phytophthora blight (*Phytophthora capsici*). *Plant health progress* 15: 19-24.
- FERREIRA, DF. 2011. Sisvar: a computer statistical analysis system. *Ciência e Agrotecnologia* 35: 1039-1042.
- GILARDI, G; BAUDINO, M; MOIZIO, M; PUGLIESE, M; GARIBALDI, A; GULLINO, ML. 2013. Integrated management of *Phytophthora capsici* on bell pepper by combining grafting and compost treatment. *Crop Protection* 53: 13-19.
- GRANKE, LL; QUESADA-OCAMPO, L; LAMOUR, KH; HAUSBECK, MK. 2012. Advances in research on *Phytophthora capsici* on vegetable crops in the United States. *Plant Disease* 96: 1588-1600.
- HONG, JC; NORMAN, DJ; REED, DL; MOMOL, MT; JONES, JB. 2012. Diversity among *Ralstonia solanacearum* strains isolated from the southeastern United States. *Phytopathology* 102: 924-936.
- JARQUE, C; BERA, A. 1987. A test for normality of observations and regression residuals.

- International Statistical Review* 55: 163-172.
- LEBEAU, A; DAUNAY, MC; FRARY, A; PALLOIX, A; WANG, JF; DINTINGER, J; CHIROLEU, F; WICKER, E; PRIOR, P. 2011. Bacterial wilt resistance in tomato, pepper, and eggplant: Genetic resources respond to diverse strains in the *Ralstonia solanacearum* species complex. *Phytopathology* 101: 154-165.
- LOPES, CA; BOITEUX, LS. 2004. Biovar-specific and broad-spectrum sources of resistance to bacterial wilt (*Ralstonia solanacearum*) in *Capsicum*. *Crop Breeding and Applied Biotechnology* 4: 350-355.
- LOPES, CA; BOITEUX, LS; ESCHEMBACK, V. 2015. Eficácia relativa de porta-enxertos comerciais de tomateiro no controle da murcha-bacteriana. *Horticultura Brasileira* 33: 125-130.
- MADEIRA, NR; AMARO, GB; MELO, RAC; RIBEIRO, CSC; REIFSCHNEIDER, FJB. 2016. Compatibilidade de porta-enxertos para pimentão em cultivo protegido. *Horticultura Brasileira* 34: 470-474.
- NASS, LL; SOUZA, KRR; RIBEIRO, CSC; REIFSCHNEIDER, FJB. 2015. Synthesis of a base population of Habanero pepper. *Horticultura Brasileira* 33: 530-532.
- NARESH, P; MEENU, K; ACHARYA, GC; REDDY, AC; LAKSHMANA, DCR. 2019. Genetics and molecular markers for resistance to major soil borne pathogens in chilli (*Capsicum annum* L.). *Research Journal of Biotechnology* 14: 101-105.
- PETRY, R; PAZ-LIMA, ML; BOITEUX, LS; CAFÉ-FILHO, AC; REIS, A. 2016. Reaction of *Solanum* (section *Lycopersicon*) germplasm to *Phytophthora capsici*. *European Journal of Plant Pathology* 148: 481-489.
- PINHEIRO, JB; REIFSCHNEIDER, FJB; PEREIRA, RB; MOITA, AW. 2014. Reação de genótipos de *Capsicum* ao nematoide das galha. *Horticultura Brasileira* 32: 371-375.
- REEVES, G; MONROY-BARBOSA, A; BOSLAND, PW. 2013. A Novel *Capsicum* gene inhibits host-specific disease resistance to *Phytophthora capsici*. *Phytopathology* 103: 472-478.
- RIBEIRO, CSC; BOSLAND, PW. 2012. Physiological race characterization of *Phytophthora capsici* isolates from several host plant species in Brazil using New Mexico recombinant inbred lines of *Capsicum annum* at two inoculum levels. *Journal of the American Society for Horticultural Science* 137: 421-426.
- RIBEIRO, CSC; SOUZA, KRR; CARVALHO, SIC; REIFSCHNEIDER, FJB. 2015. BRS Juruti: the first Brazilian habanero-type hot pepper cultivar. *Horticultura Brasileira* 33: 527-529.
- ROSSATO, M; SANTIAGO, TR; LOPES, CA. 2018. Reaction of *Capsicum* peppers commercialized in the Federal District to bacterial wilt. *Horticultura Brasileira* 36: 173-177.
- SÁNCHEZ-CHÁVEZ, E; SILVA-ROJAS, HV; LEYVA-MIR, G; VILLARREAL-GUERRERO, F; JIMÉNEZ-CASTRO, JA; MOLINA-GAYOSSO, E; GARDEA-BÉJAR, AA; ÁVILA-QUEZADA, GD. 2017. An effective strategy to reduce the incidence of *Phytophthora* root and crown rot in bell pepper. *Interciencia* 42: 229-235.
- SHANER, G; FINNEY, RE. 1977. The effect of nitrogen fertilization on the expression of slow-mildewing resistance in knox wheat. *Phytopathology* 67: 1051-1056.
- STEINER, OSR; BOSLAND, PW. 2008. Recombinant inbred line differential identifies race-specific resistance to *Phytophthora* root rot in *Capsicum annum*. *Phytopathology* 98: 867-870.
- STEEL, RGD; TORRIE, JH; DICKEY, DA. 1997. Principles and procedures of statistics: a biometrical approach. 3.ed. New York: McGraw Hill Book. 666p.
- TEODORO, AFP; ALVES, RBN; RIBEIRO, LB; REIS, K; REIFSCHNEIDER, FJB; FONSECA, MEN; SILVA, JP; AGOSTINI-COSTA, TS. 2013. Vitamin C content in Habanero pepper accessions (*Capsicum chinense*). *Horticultura Brasileira* 31: 59-62.
- VÁZQUEZ, PV; CORTEZ, MCN. 2018. Use of organic alternatives in the production system of habanero pepper (*Capsicum chinense* Jacq.) under greenhouse conditions. *African Journal of Agricultural Research* 13: 1091-1094.
- WICKER, E; GRASSART, L; CORANSON-BEAUDU, R; MIAN, D; GUILBAUD, C; FEGAN, M. 2007. *Ralstonia solanacearum* strains from Martinique (French west indies) exhibiting a new pathogenic potential. *Applied and Environmental Microbiology* 73: 6790-6801.
- WINSTEAD, NN; KELMAN, A. 1952. Inoculation techniques for evaluation of resistance to *Pseudomonas solanacearum*. *Phytopathology* 42: 628-634.